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Nest building and description of parental care behavior in a temperate reef fish, *Chromis crusma* (Pisces: Pomacentridae)

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Abstract

Background: Subtle ecological and behavioral mechanisms that enhance reproduction such as nest building by animals may provide useful information of population level processes. Variation in behavioral traits may be observed as phenotypic traits that are shaped by sexual and natural selection. Using ecological sampling of benthic substrata and underwater video recordings, we assessed nest-building behavior through habitat modifications, and size of individuals performing parental care behavior and egg/hatching traits of an abundant temperate reef fish, *Chromis crusma*.

Results: We identified that only male individuals performed nest building and uniparental care. We noted that nests containing filamentous green or red algae had the highest percent cover of eggs. Using video recordings, we provided evidence of parental care. Male individuals spent nearly 80% of their time inside the nest, aerating the eggs with their fins and mouth, removing unwanted materials, and defending the nest against conspecific, heterospecific, and other predators. The field-collected eggs, characterized by an oil globule and adhesive filaments, hatched after 7 days in the laboratory. The nest-building and parental care behavior of *C. crusma* lasted for 3 months, and several nests can be constructed throughout the season.

Conclusion: The behavior of building and defending the nests, which is a characteristic of the family members, is a key aspect for the success of the *C. crusma*; this fish is abundant in kelp ecosystems of the southeastern Pacific.

Keywords: Reproductive strategies; Parental care; Reef fishes; Chile

Background

Plants and animals have evolved to ensure the survival of their own future genes, thus affecting the number of offspring for future generations (Gross 2005). Furthermore, natural selection is expected to favor behaviors that maximize lifetime reproductive success where mixtures of behavioral phenotypes may exist among species and populations (Alcock 2009). Individuals must therefore perform a series of behaviors to obtain their optimum reproduction over the course of a lifetime (Darwin 1859). Teleost fish display a wide range of parental care behavior, ranging from no care at all, uniparental care by a female or male, to biparental care (Gross and Sargent 1985; Blumer 1982; Webb et al. 1999). In order to succeed in their offspring's survival, individuals need to evaluate the benefits of conducting care, resulting in variation or termination of parental care behavior within populations (Ochi 1985; Smith and Wooton 1995).

The numerous costs associated to finding a mate, competing with conspecifics, courtship, and parental care are often assessed by individuals, allowing behavioral adjustments to minimize these costs (Itzkowitz 1990; Clotfelter et al. 2006). Parental care activities occur after fertilization. In fish, a common care activity is fanning, in which the parent moves the pectoral, anal, and/or pelvic fins over the egg clutch, eliminating sediments from the surrounding environment (Goulet 1995). More sophisticated parenting methods involve the movement of the mouth or branchial cavity, which in turn oxygenates the eggs and removes suspended sediments, or the direct, selective removal of damaged or dead eggs with the mouth (Goulet 1998; Ochi 1985). Chasing competitors and predators away from the nest are also important forms of



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parental care (Amundsen 2003). Egg characteristics may also be associated with adult behavior. For example, egg size and development time can depend on predominance and duration of parental care (Sargent et al. 1987; Shine 1978).

The most noticeable behaviors of several fish species for attracting mates are resource and mate monopolization (Blumer 1982; DeMartini and Sikkel 2006; Baylis 1981). Territorial fishes, for instance, can acquire mates by establishing a temporary breeding territory or nesting site through the formation of structures used to house eggs (Sikkel 1995; Rushbrook et al. 2008; Goulet 1998; Kawase et al. 2013). Among fishes, nest construction can be divided into three categories: a) excavations - burrows in gravel, sand, or mud (Kawase et al. 2013); b) simple walls piling up sand, pebbles and shells, and artificial substrata (Colin 1973); and c) cement - secreting chemicals from the kidneys to glue together algae, detritus, or sand (Rushbrook et al. 2008).

In marine benthic substrata, nest builders clean their nesting areas by removing and/or farming preferred substratum species in an area where eggs have been deposited (Unger and Sargent 1988; Klumpp and Polunin 1989). Nests can be pivotal for courtship because they may attract the opposite sex and provide shelter for the developing eggs (DeMartini and Sikkel 2006). Nest quality is expected to differ among individuals who construct them (Clotfelter et al. 2006). In marine environments, individuals may face several limitations in building nests such as a) bottom topography (e.g., vertical walls versus horizontal surfaces), b) abundance of preferred nest material, c) presence of heterosexual or conspecific breeding pairs (i.e., operational sex ratio), and d) ability to chase predators and competitors away from the nest (Coleman and Wilson 1998; Taborsky 1994).

Fishes from the family Pomacentridae are oviparous (DeMartini and Sikkel 2006), and there are substantial differences in the mating systems within members of this family in both temperate and tropical systems (see Table 1). Nest building along with parental care behavior has been observed in 25 species, 7 of which are temperate (Table 1). The male is responsible for parental care and nest building in 80% of the cases and biparental care in the remaining 20%. Biparental care is present for tropical species only. The majority of the species that build nest do not discriminate the species in the substrata to lay their eggs. However, algal species (usually turf algae) was the preferred site for spawning for six species, three of them are temperate (including Chromis crusma) and two subtropical species. Artificial structures, dead coral, and shells are the preferred sites for six tropical species. Most tropical species deposit their eggs under crevices or rock boulders (Table 1).

Castañeta, *C. crusma*, is a common and widely distributed damselfish inhabiting the Southeast Pacific Ocean from

Santa Rosa Island (5° SL), Ecuador, to Valdivia (39° SL), Chile (Pequeño et al. 2005). Along the central Chilean coast, individuals are usually found in rock outcrops within depths of between 5 and 35 m and reef areas dominated by large brown macroalgae where they recruit (i.e., *Lessonia trabeculata*). Adults can reach up to 25 cm in total length (TL) and are often seen in large schools (up to 100 individuals) (Pérez-Matus et al. 2007). The diet of *C. crusma* is comprised mainly of planktonic species (Angel and Ojeda 2001). As a first step to understand variability in parental care and nest-building behavior of an abundant temperate reef fish, we describe the information about the nest and the type of parental care and correlated this behavior with descriptions of the benthic eggs and recently hatched larvae.

Methods

Study site

Field observations on nesting behavior were performed during 2011 to 2012 and nest-building behavior was observed during November 2011 through March 2012, which coincides with the spawning season of *C. crusma* at Zapallar, Valparaíso (32° 33′ SL to 71° 28′ WL) (Figure 1). We searched for and surveyed nests along the depth distribution of *C. crusma*, within depths of between 5 and 35 m. Large boulders covered with turf and calcareous crustose algae dominated the shallow sections of the study site, where kelp was sparsely distributed from 5 to 15 m. Encrusting sessile fauna such as sponges (*Halichondria* sp. and *Clionopsis platei*) and sea anemones (*Parazoanthus* sp. and *Anemonia alicemartinae*) as well as filamentous green and red algae dominated the substratum from 15 to 30 m (Figure 1a,b).

Habitat modifications by C. crusma

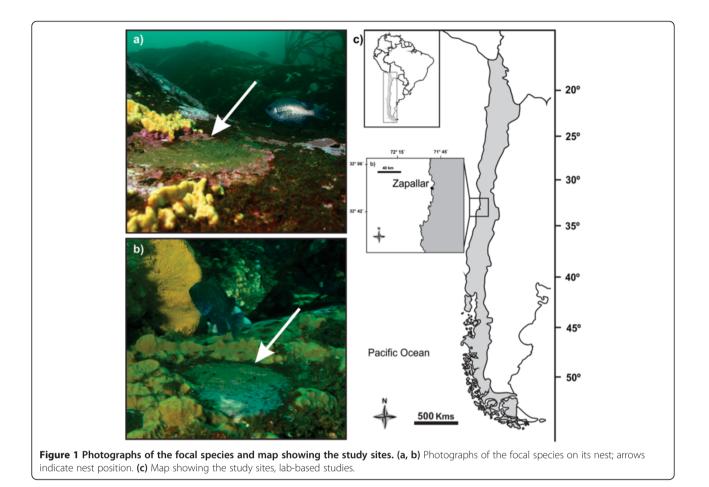
We searched for demersal nests of C. crusma, defining a nest as an area containing eggs attached to a substratum that was guarded by an individual (Figure 1a,b). At each nest, we described the color of the fish and estimated the size of the nest defender using a 4-cm dive weight positioned 5 cm from the nest. Mean nest size (cm) was calculated by averaging the maximum horizontal and vertical lengths of the nest measured to the nearest 0.5 cm using a measuring tape. We categorized the substratum slope where the nest was located as either a) vertical walls, b) horizontal platforms, or c) intermediate slope platforms. Differences in nest size among the substratum slope were tested using a one-way ANOVA. To meet ANOVA assumptions of normality and homoscedasticity of variance, we used a Cochran *c*-test and Fligner-Killeen test, respectively (Crawley 2007). To confirm our prior field observations that nest defenders were males, we randomly collected six adult individuals from six different guarded nests and determined the sex of each individual.

Species	Distribution	Lt	Type of nest	Egg	Sex	Reference
Abudefduf abdominalis	Tropical: Eastern Central Pacific	30	Substratum	7	Male	Ridley 1978
Abudefduf luridus	Temperate: Eastern Atlantic	15	Vertical walls, algae, and sponges	ND	Male	Ridley 1978; Mapstone and Wood 1975
Abudefduf saxatalis	Temperate: Atlantic Ocean	23	Reef wall, crevices	5	Male	Reese 1964
Abudefduf taurus	Tropical: Western Atlantic	25	Crevices	7	Male	Fishelson 1998; Mapstone and Wood 1975
Abudefduf zonatus	Tropical: Indo-Pacific	13	Sand, under rock, and coral rubble	ND	Male	Ridley 1978; Mapstone and Wood 1975
Acanthochromis polyacanthus	Tropical: Western Pacific	14	Dead coral under rocks	30	Biparental	Ridley 1978; Robertson 1973
Amblyglyphidodon leucogaster	Tropical: Indo-West Pacific	13	Dead coral or artificial structures	8	Male	Goulet 1995, 1998
Amphiprion bicinctus	Tropical: Western Indian Ocean	14	Substratum	6	Biparental	Gittleman 1981
Amphiprion melanopus	Tropical: Pacific Ocean	12	Substratum	9	Biparental	Breder and Rosen 1966; Gittleman 1981
Amphiprion percula	Tropical: Western Pacific	11	Substratum	6	Biparental	Ridley 1978
Amphiprion xanthurus	Tropical: Indo-West Pacific	15	Substratum	ND	Biparental	Gittleman 1981
Chromis caeruleus	Temperate: Mediterranean Sea	9.5	Green and turf algae	4	Male	Sale 1971
Chromis chromis	Tropical and temperate: Eastern Atlantic and Mediterranean Sea	25	Under crevices	3	Male	Ridley 1978; Mapstone and Wood 1975
Chromis cyanea	Tropical: Western Atlantic and the Caribbean Sea	15	Under crevices	4	Male	Albrecht 1969; Ridley 1978
Chromis dispilus	Temperate: Southwest Pacific (New Zealand)	21	Substratum	6	Male	Kingsford 1985
Chromis multilineata	Tropical: Western Atlantic and the Caribbean Sea	20	Under crevices, in Sargassum sp.	3	Male	Albrecht 1969; Myrberg et al. 1967
Chromis notata	Temperate: southern Japan, Ryukyu Islands, Taiwan, and China	17	Substratum	7 ^a	Male	Ochi 1985
Chromis punctipinnis	Temperate: Eastern Pacific	25	Clean rock ledges or small caves	8	Male	Limbaugh 1964; Turner and Ebert 1962
Dascyllus aruanus	Tropical and temperate: Indo-West Pacific and north to southern Japan, south to Sydney, Australia	10	Substratum	3	Male	Mizushima et al. 2000
Stegastes leucorus	Tropical: Eastern Central Pacific	20	Substratum	3 to 5	Male	Ridley 1978
Stegastes leucostictus	Tropical: Western Atlantic	10	Shells under crevices	3 to 5	Male	Ridley 1978
Stegastes partitus	Tropical: Western Atlantic and the Caribbean	10	Monolayer of species	3 to 5	Male	Knapp and Warner 1991
Hypsypops rubicunda	Temperate and tropical: Eastern Central Pacific and northern central Baja California	30	Red algae	3	Male	Clarke 1970
Pomacentrus amboinensis	Tropical: Western Pacific	15	Dead coral	4	Male	Murphy et al. 2007; Maddams and McCormick 2012
Pomacentrus negasakiensis	Tropical: Indo-West Pacific	11	Boulders, artificial substrata	4	Male	Moyer 1975
Premnas biaculeatus	Tropical: Indo-West Pacific	17	Substratum	7	Biparental	Fishelson 1998; Ridley 1978

Table 1 Review of type of nest among tropical and temperate pomacentrid members

Geographic distribution, total length of individuals (Lt, cm), egg development in days (Egg), and sex of the individuals who perform care to their young. ND refers to not determined parameter. ^aMean number of days from 30 April until 24 August 1982 (Ochi 1985).

To describe nest habitat and substrate composition around the nest, we selected nests (n = 40) that were guarded by solitary individuals, and three quadrats were each randomly placed inside the nests, at the edge of the nests (along a boundary of the egg clutch), and outside the nests (where eggs are absent). In each quadrat, we recorded the percent cover of sessile benthic species using a 0.01-m² quadrat with 15 intersection points (random point contact,



RPC). We quantified the percent cover of eggs inside the nest. When necessary for confirmation of species identity, eggs and all sessile species were collected by manually removing with a spatula and preserved in 5% buffered formalin. We also used underwater photography (Nikon-D90 with Ikelite Housing and two D125 substrobe; Ikelite Underwater Systems, Indianopolis, IN, USA) on quadrats to further analyze the images on a computer. The same divers performed all sampling by means of self contained underwater breathing apparatus (SCUBA) throughout the study.

To determine differences in the substrata composition inside and outside of nests, as well as among nests, we used multivariate analyses. A Bray-Curtis resemblance matrix was used to carry out the analysis, and a one-factor permutational multivariate analysis of variance (PERMANOVA; Anderson 2001) was used to test for significant differences in the substratum composition within and among the nests. At each spatial scale, the substratum species compositions were analyzed using similarity percentages (SIMPER), identifying the species that contributed most to the differences between the nest scales (i.e., outside, edge, inside). We estimated the nest quality by evaluating the relationship between species richness (i.e., benthic composition) and the number (i.e., percent cover) of eggs in each nest (Sikkel 1995).

Parental care: nest-guarding behavior

In order to avoid diver effects on the nest-guarding behavior (individuals tended to leave the nest when divers approached), we set up an underwater video camera (Go-Pro[™] Hero 2, GoPro, Inc., San Mateo, CA, USA) anchored with a 2-kg weight. The total dimension of the camera was 8 × 8 cm. The camera was randomly positioned 30 cm from the nests, and the recording time ranged from 25 to 35 min. We placed the camera in eight nests totaling approximately 280 min of recording time. Filming occurred during early egg development stages (2 to 3 days). Images were downloaded to a computer, and we quantified the time that each recorded individual spent performing the following: a) time inside the nest, b) aerating the eggs with the mouth, c) fanning with pectoral or anal fins, d) removing unwanted drift material, and e) defending the nest against intruders.

Embryonic development and larval growth

In order to describe the development time, hatching, and larvae growth of *C. crusma*, we collected eggs (>50) from more than ten nests at different depths using SCUBA and a scraping knife. The eggs were carefully placed in a 1-L glass container and transported in glass jars with seawater

and ice packs to the laboratory (32°56′ SL, 71°33′ WL) (Figure 1c). Once in the laboratory, egg clutches were kept aerated with filtered seawater (0.5 μ m) inside 5-L glass jars at 18°C. Embryo development was evaluated on a daily basis until hatching occurred (see below). Recently hatched larvae were transferred into 1-L glass jars at a density of 30 larvae L⁻¹. When the yolk sac was depleted, larvae were fed with *Nannochloropsis* spp. and rotifers *Brachionus plicatilis*.

During each monitoring, all eggs and larvae were photographed with a stereomicroscope Olympus SZ-61 equipped with a Motic video camera 2500 (5.0 MP; Olympus Corporation, Tokyo, Japan) connected to a PC with Moticam Image Plus 2.0 software. All measurements (mm) were carried out on fresh eggs (length, maximum width, oil globule diameter) and larvae [body length (from the tip of the snout to the tip of the notochord), pre-anal length, eye diameter, yolk sac length and height, and oil globule diameter]. Egg and yolk sac volume were estimated considering the yolk sac as an ellipsoid $(V = 4/3\pi ab^2)$, where a is half of the egg or yolk sac length (mm) and b is half of the height (mm) (Bustos and Silva 2011). Larval growth was estimated adjusting a least squares linear regression model between body length (BL, mm) and age (days post hatch, dph), $BL = \alpha + dph\beta$, where α and β are the size at hatching and larval growth rate respectively, estimated by the model. All statistical tests were performed using R v 2.14 (R Development Core Team 2012).

Results

Habitat modifications of *C. crusma* during the spawning season

According to our observations, reproductive behavior in C. crusma commenced in late October and ended in March (austral spring-summer). During this period, we sampled every 2 weeks, a total of six different sampling days. More than 80 nests were observed, and of these, a total of 39 nests were measured, with a mean diameter of 23.5 cm (\pm 5.2 SD). All adult individuals collected (n = 6) from the nests were males. All had dark grey coloring that differed from their pelagic counterparts and other benthic transients, who were pale grey in color. Our visual estimates revealed that nest defenders ranged from 15.0 to 21.5 cm TL, with a mean of 17.3 cm TL (\pm 2.6 SD). There was no relationship between fish TL and nest diameter (linear regression $r^2 = 0.08$, df = 1, *F* = 0.09, *p* value >0.05). Mean nest size measured 21.3 (±2.2 SE), 23.3 (±1.3 SE), and 24.6 (±1.5 SE) cm in vertical walls, intermediate slope, and horizontal platforms, respectively. No significant differences were observed in the size of the nest with respect to the substratum slope (one-way ANOVA, df = 2; *F* = 0.4, *p* value >0.05).

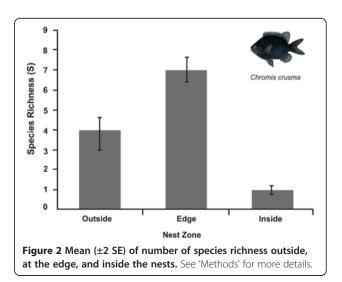
In the study area, we recorded a total of 16 benthic species that covered the substratum outside, inside, and

at the edge of the nests. Significant differences were found among these categories (one-way ANOVA, df = 2; F = 6.5, p value <0.01). We detected more benchic species at the edge and outside of nests than inside them (Tukey post hoc, p value <0.01, Figure 2). Moreover, outside and edge richness varied significantly among nests, mirroring local changes in the surrounding dominant community structure (i.e., sessile animal or algae-dominated substratum). In contrast, species richness inside of nests was low and similar among nests regardless of the surrounding community. In terms of nest quality, a significant negative relationship was observed between the percent cover of eggs and the number of benthic species (linear regression, $r^2 = 0.4$, df = 1, F = 7.1, p value <0.001). A high number of eggs were present in nests that contained a small number of species (Figure 3).

Defended nest habitats were composed predominantly of filamentous green algae (mostly *Cladophoropsis herpestica*) or red algae (mostly *Pterosiphonia dendroidea* or *Schottera nicaeensis*) and/or crustose algae (*Lithothamnion* sp.) (Figure 4). The benthic sessile community composition differed inside the nests compared with the species that covered the edge of the nest or outside of them (PERMA-NOVA, df = 2, pseudo F = 3.8, p value <0.01). Cumulative contributions (SIMPER) of the most influential species indicated that green filamentous algae accounted for 54% and 56% of the differences between inside the nest and outside or edge of the nest, respectively. The percentage cover of these algae was higher inside the nests (Figure 4).

Parental care: nest-guarding behavior

The video recordings of *C. crusma* (see video, http:// vimeo.com/89531693) revealed that males spent between 80% and 95% of their total time inside rather than outside the nests. Fanning was the most frequent activity performed by *C. crusma* inside the nests, ranging from



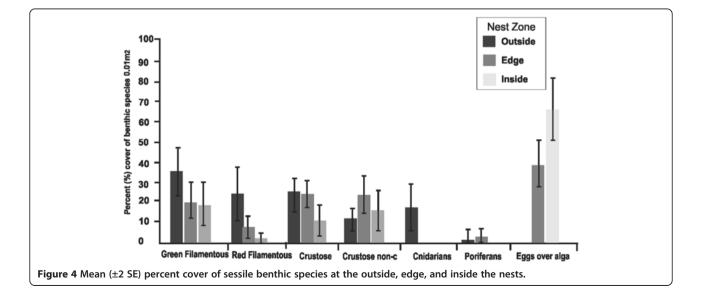
78% to 94% of the time. Defending the nest was the second most important activity (7% to 15% of the time). Predators such as the sandperch (Pinguipes chilensis), the triplefin (Helcogrammoides spp), and the rock shrimp (Rhinchocynetes typus) were excluded from the nest by exaggerated swims and accelerated chasing movements. The display of dorsal and pectoral fins was also an aggressive behavior performed by defending males. Only during short (few seconds), aggressive chases to ward-off potential predators and conspecifics would defending males leave the nest. In addition to protective behaviors, males were observed to actively aerate eggs by removing individuals and holding them within an open mouth faced directly in the direction of water flow for several seconds before placing them back in the nest. This was a commonly observed behavior (average of 24.5 events per individual) for the majority (95%) of video/recorded individuals. Finally, the removal of drift material such as dead eggs and other detritus that can affect the normal development of the eggs was also observed.

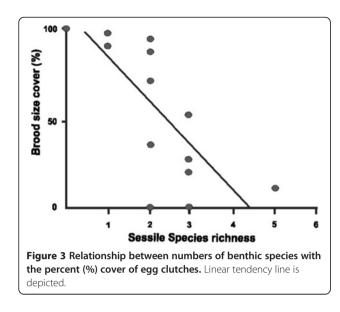
Embryonic development and larval growth

The demersal eggs of C. crusma were ovoid to elliptical and ranged in size from 0.96 to 1.20 mm (mean: 1.08 mm ± [0.06 SD], n = 50) on the long axis and 0.65 to 0.72 mm (mean: 0.68 ± 0.02 mm SD) in maximum width. Estimated volume ranged from 0.23 to 0.30 mm^3 (mean: 0.2 ± 0.02 mm³ SD). Eggs contained one single red to yellow oil globule, 0.18 to 0.31 mm in diameter $(0.25 \pm 0.02 \text{ mm SD})$. Adhesive filaments attached eggs from their basal poles to the nest (Figure 5). Embryonic development lasted 7 to 8 days, and hatched larvae measured between 2.85 and 3.67 mm in body length (mean 3.30 mm \pm 0.18 SD), with a yolk sac of $0.021 \pm 0.004 \text{ mm}^3$ (range: 0.013 to 0.030 mm³) that was generally depleted after 2 dph. The oil globule was completely absorbed after 7 dph. Both estimated slope (growth rate) and intercept (hatch size) were significantly different from zero (p value_{β} = 0.0014; p value_{α} <0.001).

Discussion

Our results revealed that male individuals of *C. crusma* set up nesting territories during the austral summer (November through March), which they aggressively defend and actively clean/maintain. Moreover, males frequently undertook numerous 'incubating' activities that commonly benefit egg development, including aeration via fin fanning and exposing eggs to direct water flow, as well as egg re-positioning within the nest. The negative correlation between egg abundance and benthic species richness conforming nest substrate suggests that important differences in nest quality occur within a site, which







was unrelated with the size (i.e., total length) of the individuals performing the care.

Male individuals responsible for the fate of the fertilized eggs and success during development result in uniparental care (Gross 2005). Males defend the nest against predators, aerate and fan the eggs, and remove drift materials from the nest during eight consecutive days after which larvae hatch in the laboratory at 18° C. Finally, one oil globule and adhesive filaments characterize *C. crusma* eggs and many other members of the genera (see Kingsford 1985), which may explain the upkeep of benthic filamentous algae inside the nest by male individuals (as it allows egg attachment to the surface).

Methods of nest construction by *C. crusma* coincide with other temperate reef fishes of the family Pomacentridae (see Table 1). For example, the temperate damselfish *Hypsypops rubicundus* exhibits a similar behavior in the preparation of the nesting site, maintaining a red alga in the middle of the cleaned area, which is approximately 33 to 50 cm in diameter (Sikkel 1995). In general, temperate reef fish use benthic algae for depositing the eggs while tropical members use dead coral or artificial materials. Tropical members deposit eggs under crevices and boulders presumably to avoid egg predators, which are potentially more abundant in tropical reefs than predators in temperate reefs (Table 1).

Nest construction, in turn, can be crucial to increasing the reproductive success of male individuals. There is striking recent evidence, which suggests that nest and nest-building behavior have evolved under sexual selection (Borgia 1985; Tomás et al. 2013; Kawase et al. 2013). Nevertheless, a problem that has been puzzling behavioral ecologists is the identification of those aspects of the nest that influence the success of the spawning male, and thus determine what affects females in their selection of the spawning site (but see Tomás et al. 2013). In C. crusma, the identification of nest features that influence female choice can be complicated as is the case with many other fishes which perform parental care; females in many populations preferably spawn in nests that already contain eggs (Hoelzer 1990). Sex dissimilarities in shape, color, and behavior result from differences in the reproductive achievement of individuals (Clotfelter et al. 2006). Future research should be directed towards understanding the role of nest features as sexual signals (see Kawase et al. 2013).

A correlation between benthic sessile species and cover of eggs (as proxy of brood size) was observed in this study. It has been reported that size of the brood is crucial to the survival of offspring (see Ochi 1985). Sexual selection forces may well fit into this, as male individuals may have acquired information from past investments in nest building by carefully selecting sites where monospecific filamentous algae thrive. Alternatively, males can clean the substratum in such sites maintaining the preferred algae for females to deposit their eggs. The type of benthic substrata (not the bed-rock slope) may also be important for female choice in this species.

Considerable debate exists in the literature surrounding which hypothesis came first, the evolution of egg size (small vs large eggs) or the evolution of parental care. Shine (1978) proposed the 'safe harbor' hypothesis that predicts the evolution of parental care prior to the evolution of large egg size (selection favors the increase in egg size). Nussbaum and Schultz (1989) challenged this by stating that a form of care should be incorporated to reduce the long-term developmental mortality in relation to egg clutch size. Our study is not attempting to resolve this conflict. The high number of egg predators may indicate the high energetic lipid value and volume of C. crusma eggs (an oil globule), which correlates with the amount of energy spent by adults in maintaining predator distance. Differences in egg developmental time exist among tropical and temperate pomacentrids (Table 1). Egg development is similar among temperate reef fish, with time from egg deposit to hatching lasting 6 to 8 days for most temperate species and among tropical pomacentrids with 3 to 5 days for most tropical and subtropical pomacentrids. Some notable exceptions were present in tropical species such as Acanthochromis leucogaster, which exhibited extended egg development time of up to 30 days (Table 1). The longer time for egg development in temperate reef fish may directly affect the cost of parental care of individuals.

The complex life cycle (pelagic larval phase, benthic juvenile and adult) of most marine fishes is characterized by the high mortality of eggs and larvae. A bottleneck of population structure is often observed at the egg and larval stage. Therefore, nest construction and parental care may have important evolutionary consequences that affect local population structures (see Reynolds et al. 2002 for review). Temperate reef fish exhibit a diverse array of reproductive modes, life histories, and parental care systems. With substantial variation among individuals, the reproductive behavior of pomacentrid fishes can be divided into the establishment of a breeding territory, nesting site, construction of nests by building structures used to house eggs, courtship and pair formation, spawning and fertilization, and finally parental care (Limbaugh 1964; Clarke 1970; Sikkel 1995; Turner and Ebert 1962). To our knowledge, this is the first study which describes parental care carried out by a pomacentrid temperate reef fish in the southeastern Pacific. C. crusma is abundant within the reef system (Pérez-Matus et al. 2007). Modes of parental care have co-evolved in many taxa, and comparative information about young survivorship from different modes of reproduction is needed. Environmental and demographic parameters influencing parental care patterns are promising lines for future research.

Conclusions

Our results suggest that males Chromis crusma created their nest by removing some species from the substrate, selecting only sessile algae species for egg deposition. The number of sessile species removed may provide insights to the nest quality, which is unrelated to male's size. As in many other damselfish, egg development is longer in temperate that in tropical species. In laboratory, egg development lasted 7-8 days, which coincides with the time spent by individuals in defending their nest. Males aggressively defended their nest against predators, maintaining eggs by constantly aerating and cleaning eggs by removing drift material and repositioning eggs within the nest. Although this research provides a first step into the understanding of the biology and reproductive behavior of an abundant reef fish of the southeastern Pacific, environmental and demographic (i.e., operational sex ratio) parameters affecting this behavior requires further research.

Competing interests

The authors declare that they have no competing interests.

Authors' contribution

TNF and APM conceived the study, participated in the design and coordination of the manuscript, and carried out the field sampling. ML and CB carried out the larval description, development, analysis, and the written section. APM and TNF wrote the paper. All authors read and approved the final manuscript.

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